MINOR XANTHONES OF GARCINIA MANGOSTANA

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Abstract—Two new trioxygenated xanthones with a 3,3-dimethyl allyl side chain have been isolated from the fruit hulls of *Garcinia mangostana*. The structures were established from spectral and chemical data.

INTRODUCTION

In the course of our studies on minor xanthones of Garcinia mangostana the isolation of a new 1,3,6,7-tetra oxygenated xanthone (1) has already been reported [1]. In the present paper we report the isolation and characterization of two new 1,3,5- and 1,3,7-trioxygenated xanthones which are possibly the biogenetic precursors of 1,3,6,7- and 1,3,5,8-tetraoxygenated xanthones reported earlier [1-5] from the fruit hulls of the same species. The co-occurrence of 1,3,5- and 1,3,7-trioxygenated xanthones in the same fruit hull source suggests [6] that these xanthones are derived from the same benzophenone precursor 2 by oxidative cyclization at two alternative positions.

RESULTS AND DISCUSSION

Petrol extraction of the dried and powdered fruit hulls followed by column chromatographic separation afforded two new xanthones in addition to the xanthones reported [1] earlier. The minor xanthone 3a, $C_{19}H_{18}O_5$ (M⁺ 326) showed UV and IR spectra characteristic of 1,3,5-trioxygenated xanthones [7]. The ¹H NMR spectrum of the compound in DMSO-d₆ gave a signal at δ 13.1 (1H, s) for the C-1 OH and signals at δ 5.2 (1H, t), 3.3-3.6 (2H, d), 1.76 (3H, s) and 1.66 (3H, s) confirming the presence of an isopentenyl side chain at a position other than ortho to the xanthone carbonyl. Moreover, the NMR splitting pattern of the aromatic proton signals resembled those of 1,2,3,5-tetrasubstituted xanthones [7, 8] having a downfield quartet at δ 7.65 (H-8) and a multiplet at δ 7.35 (H-6 and H-7) indicating an ABX system. The one proton singlet at δ 6.74 (H-4) and the upfield shift of the last signal in comparison with the chemical shift of the same proton of the C-3 unoxygenated xanthones suggests the presence of OMe substitution at C-3 if not OH in 3a. A positive Gibbs' test and the absence of a NaOAc induced shift in the UV also support the above OMe substitution. In the mass spectra the presence of two characteristic peaks, in addition to the M⁺, arising from the loss of 43 and 55 amu from the M⁺

IR and UV spectra of the second minor xanthone 4a, $C_{19}H_{18}O_5$ (M⁺ 326) clearly indicated it to be a 1,3,7trioxygenated xanthone [12] whereas the mass spectra showed a close similarity to the xanthone 3a suggesting the xanthone 4a, also a dihydroxy-monomethoxy-monoisopentenyl substituted xanthone where one methoxyl and one hydroxyl is ortho to the isopentenyl side chain. Moreover, the ¹H NMR spectra of the compound in DMSO- d_6 suggested that both 4a and 3a have a similar substitution pattern in ring B of the xanthone nucleus. The NMR splitting pattern of the downfield aromatic protons, which clearly differs from 3a, showed a complex 3 proton multiplet at δ 7.3-7.6 indicating a typical ABC system [13] suggesting the position of the OH substituent at C-6 or C-7. Whereas in the ¹H NMR spectra of the diacetate of 3a the 3 proton multiplet of this downfield aromatic region had been resolved to a one proton broad singlet at δ 7.95 (H-8), a two proton multiplet at δ 7.42 (H-5 and H-6) confirmed the presence of the C-7 OH. This assignment is also supported by the absence of a fused NaOAc shift in the UV spectrum. The structure 4a was finally confirmed by direct comparison of its mono-O-Me ether 4b with 1-hydroxy-3,7-dimethoxy-2-(3-methylbut-2enyl) xanthone [6].

EXPERIMENTAL

Mps are uncorr. UV spectra were in EtOH and MeOH soln. IR spectra were determined in nujol, MS were recorded at 70 eV, $^1\text{H NMR}$ spectra were taken at 90 MHz in CDCl₃/DMSO- d_6 and chemical shifts are given in δ (ppm) scale relative to TMS. $^{13}\text{C NMR}$ measurements were made with a Fourier transform

strongly suggested [9] the presence of C-1 OH and C-3 OMe groups, both ortho to the C-2 isopentenyl side chain. Furthermore, the mono-O-Me ether **3b** and the di-O-Me ether **3c** of the xanthone **3a** were found to be identical (IR, UV and MS) with the mono-O-Me and di-O-Me ether of 1,3-dihydroxy-2-(3-methylbut-2-enyl)-5-methoxy-xanthone [7,10]. Whereas the cyclodehydrogenation of the xanthone **3a** by DDQ followed by methylation afforded **5** this was different from 5-O-Me-6-desoxy-jacareubin [11], but identical with the cyclodehydrogenation product of **3b**, finally confirming the structure **3a** for the xanthone.

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accessory and signal multiplicity was determined by off resonance decoupling after proton noise decoupling. The solvent D_2O provided the lock signal, chemical shifts are given in δ (ppm) scale relative to the TMS $^{1.5}C$ signal. Chemical shifts are accurate to within ± 0.02 ppm. The xanthones after TLC were detected both by I_2 and Dragendorff's reagent.

Plant material. Fully ripe fruits of G. mangostana L. were collected in Madras (India) in July 1978.

Isolation. Hulls of 200 fruits were dried, powdered (2 kg) and extracted with petrol (bp 60–80°) for 48 hr. On concentration of the extract and after TLC (Si gel) the presence of a few more yellow coloured pigments in addition to the earlier reported mangostin and gartanin was observed. The crude extract (6 g) was chromatographed over Si gel (400 g). Fractions eluted with petrol– C_6H_6 (4:1), petrol– C_6H_6 (1:1) and with C_6H_6 were rechromatographed and further purified by prep. TLC to yield xanthone 1 (125 mg), xanthone 3a (60 mg) along with xanthone 4a (25 mg) and mangostin (2 g) from the above 3 fractions, respectively.

Xanthone 1. Recrystallization from C_6H_6 as bright yellow fine crystals (100 mg), mp 152–154°; UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (log ϵ) 238 (3.95), 286 (4.51). 333 (4.11), 357 (4.16); $\lambda_{\text{max}}^{\text{MeOH}-\text{NaOAc}}$ nm $(\log \varepsilon) 238 (4.27), 274 (4.54), 300 (4.32), 360 (4.49); IR v_{max}^{nujol} cm^{-1}$ 3500 (OH), 1650 (7-pyrone C=O); MS m/e (rel. int.); 408 $(M^+, 40)$, 393 $(M^+ - 15)$, 365 $(M^+ - 43, 30\frac{0}{20})$; ¹H NMR (80 MHz, CDCl₃): δ 13.55 (1H, s), 6.73 (1H, s), 6.64 (1H, d, $J = 10 \,\text{Hz}$), 6.27 (1H, s), 6.15 (1H, s), 5.47 (1H, d. J = 10 Hz), 5.18 (1H, t, J = 6.5 Hz), 4.01 (2H, d, J = 6.5 Hz), 3.73 (3H, s), 1.76 (3H, s), 1.62 (3H, s), 1.4 (6H, s); ¹³CNMR (15.1 MHz, CDCl₃); δ 18.1 (C-4'), 25.6 (C-3 CH₃), 26.5 (C-1'). 28.3 (C-2 CH₃), 61.8 (C-8 OCH₃), 77.8 (C-2), 94.0 (C-12), 101.6 (C-10), 103.6 (C-5a), 104.4 (C-4a), 112.1 (C-6a), 115.6 (C-4), 123.1 (C-2'), 126.9 (C-3), 131.8 (C-3'), 136.9 (C-7), 142.7 (C-8), 154.5 (C-9), 155.6 (C-10a), 156.1 (C-11a), 157.8 (C-5), 159.8 (C-12a), 181.8 (C-6). (Found: C, 71.0; H, 5.6 $^{\circ}_{\circ}$, $C_{24}H_{24}O_6$ requires: C, 70.6; H, 5.9 ° a). The diacetate crystallized from MeOH as white microcrystals, mp 171–172°; MS m/e (rel. int.): 492 (M⁺, 20), 450 $(M^+ - 42, 30), 408 (M^+ - 2 \times 42, 10).$

Xanthone 3a. The solid from prep. TLC on recrystallization from MeOH afforded yellow flakes (60 mg), mp 242-244°, UV

 $L_{\rm max}^{\rm EOH}$ nm (log ε), 245 (4.53), 256 (4.48), 313 (4.21), 375 sh (3.51); IR $L_{\rm max}^{\rm rujoi}$ cm⁻¹: 3350 (chelated OH), 1650 (γ-pyrone C = O); MS m/e (rel. int.): 326 (M⁻¹, 27), 311 (M⁺ – 15, 25), 283 (M⁻¹ – 43.63), 271 (M⁺ – 55, 100); ¹H NMR (90 MHz, DMSO- L_0): 13.10 (1H, s), 7.65 (1H, q), 7.35 (2H, m), 6.75 (1H, s), 5.2 (1H, t, L_0) = 6.5 Hz), 3.99 (3H, s), 3.2–3.6 (2H, masked by L_0)–DMSO signals), 1.76 (3H, s), and 1.66 (3H, s). (Found: C, 70.1; H, 5.8%, L_0)- L_0 0, L_0 1 = L_0 1 = L_0 2 = L_0 3 = L_0 3 = L_0 4. (69.9; H, 5.5%). The diacetate prepared in the usual way was crystallized from petrol– L_0 4 as white needles, mp 153–154°, MS L_0 6 = L_0 7 = L_0 8 = L_0 9 = L

Mono-O-Me ether of 3a. Xanthone 3a (10 mg) on methylation with CH_2N_2 – Et_2O afforded 3b as yellow plates from MeOH, mp 168– 169° (lit. [7], mp 172– 173°); MS m/e (rel. int.): 340 (M $^+$, 40).

Di-O-Me ether of 3a. Xanthone 3a (10 mg) on methylation with $\rm K_2CO_3/MeI$ in $\rm Me_2CO$ afforded 3c as white fine needles from EtOH, mp 158–159° (lit. [7], mp 162–163°); MS $\it m/e$ (rel. int.) 354 (M $^+$, 60) and was found to be identical with 1,3,5-trimethoxy-2-(methylbut-2-enyl) xanthone by mmp, co-TLC and superimposable IR.

Cyclodehydrogenation of 3h and 3a. Xanthone 3b (20 mg) in C_6H_6 (10 ml) was refluxed with DDQ (10 mg) for 3 hr. The reaction mixture was filtered hot and the filtrate after evaporation and chromatographic purification afforded 5 as fine yellow crystals from C_6H_6 (16 mg), mp 226–228° (dec.). MS m/e (rel. int.) 338 (M⁺, 100), 323 (M⁺ – 15, 68), 307 (28) and 297 (30 $^{\circ\circ}_0$). Following the same method as above the xanthone 3a (10 mg) was cyclodehydrogeneated by DDQ in C_6H_6 and on methylation afforded fine yellow needles from petrol $-C_6H_6$, mp 224–227° (dec.), identical with 5 mmp, superimposable 1R.

Xanthone **4a.** On prep. TLC after purification from **3a** followed by recrystallization from C_6H_6 —MeOH, yellow fluffy crystals (25 mg), mp 220–222 °C. UV $\lambda_{\max}^{\rm EGH}$ nm (log ε) 240 (4.10), 265 (4.32), 309 (4.15) and 378 (3.90); IR $\nu_{\max}^{\rm nubol}$ cm $^{-1}$: 3250 (chelated OH), 1642 (γ-pyrone C=O); MS m/e (rel. int.) 326 (M $^+$, 20), 311 (M $^+$ – 15, 18), 283 (M $^+$ – 43, 50), 271 (M $^+$ – 55, 100), 258 (6), 241 (7 $^{\circ}$ _o); 1 H NMR (90 MHz, DMSO- d_6): 13.14 (1H, s), 7.3–7.6 (3H, m), 6.7 (1H, s), 5.16 (1H, t, J = 6.5 Hz), 3.91 (3H, s), 3.2–3.4 (2H, masked by H₂O–DMSO signals), 1.73 (3 H, s), 1.63 (3 H, s). (Found: C, 70.19; H, 5.77 $^{\circ}$ _o, $C_{19}H_{18}O_5$ requires: C, 69.9: H,

5.5%). The diacetate prepared in the usual way was crystallized from MeOH as white flakes, mp 171–172°, MS m/e (rel. int.): 410 (M⁺, 40), 368 (M⁺ – 42, 70), 326 (M⁺ – 2 × 42, 45); ¹H NMR (90 MHz, CDCl₃): 7.95 (1H, br. s), 7.42 (2H, m), 6.80 (1H, s), 5.15 (1H, t, J ~ 6 Hz), 3.96 (3H, s), 3.30 (2 H, d, J ~ 6 Hz), 2.5 (3H, s), 2.30 (3 H, s), 1.73 (3H, s), 1.55 (3H, s).

Mono-O-Me ether of 4a. Xanthone 4a (10 mg) on methylation with CH_2N_2 – Et_2O afforded 4b as yellow needles from petrol, mp 143–144° (lit. [6], mp 140°); MS m/e (rel. int.): 340 (M^+ , 45), identical with 1-hydroxy-3,7-dimethoxy-2-(3-methylbut-2-enyl) xanthone (mmp, co-TLC, and superimposable IR).

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